

This Page Is Inserted by IFW Operations
and is not a part of the Official Record

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

IMAGES ARE BEST AVAILABLE COPY.

**As rescanning documents *will not* correct images,
please do not report the images to the
Image Problem Mailbox.**

THIS PAGE BLANK (USPTO)

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : D06P 3/02, C11D 3/386	A1	(11) International Publication Number: WO 96/12846 (43) International Publication Date: 2 May 1996 (02.05.96)
<p>(21) International Application Number: PCT/DK95/00418</p> <p>(22) International Filing Date: 18 October 1995 (18.10.95)</p> <p>(30) Priority Data: 1216/94 20 October 1994 (20.10.94) DK 803/95 7 July 1995 (07.07.95) DK</p> <p>(71) Applicant (for all designated States except US): NOVO NORDISK A/S [DK/DK]; Novo Allé, DK-2880 Bagsværd (DK).</p> <p>(72) Inventors; and (75) Inventors/Applicants (for US only): PEDERSEN, Anders, Hjelholt [DK/DK]; Novo Nordisk A/S, Novo Allé, DK-2880 Bagsværd (DK). KIERULFF, Jesper, Vallentin [DK/DK]; Novo Nordisk A/S, Novo Allé, DK-2880 Bagsværd (DK).</p> <p>(74) Common Representative: NOVO NORDISK A/S; Att: Corporate Patents, Novo Allé, DK-2880 Bagsværd (DK).</p>	<p>(81) Designated States: AL, AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TT, UA, UG, US, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, MW, SD, SZ, UG).</p> <p>Published With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</p>	
<p>(54) Title: BLEACHING PROCESS COMPRISING USE OF A PHENOL OXIDIZING ENZYME, A HYDROGEN PEROXIDE SOURCE AND AN ENHANCING AGENT</p>		
<p>(57) Abstract</p> <p>The present invention relates to a process for providing a bleached look in the colour density of the surface of dyed fabric, especially cellulosic fabric such as denim, comprising use of a phenol oxidizing enzyme such as a peroxidase or a laccase, a hydrogen peroxide source and a phenothiazine or phenoxazine enhancing agent represented by formula (I).</p> <div style="text-align: center;"> <p>(I)</p> </div>		

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	GB	United Kingdom	MR	Mauritania
AL	Albania	GE	Georgia	MW	Malawi
BB	Barbados	GN	Guinea	NE	Niger
BE	Belgium	GR	Greece	NL	Netherlands
BF	Burkina Faso	HU	Hungary	NO	Norway
BG	Bulgaria	IE	Ireland	NZ	New Zealand
BJ	Benin	IT	Italy	PL	Poland
BR	Brazil	JP	Japan	PT	Portugal
BY	Belarus	KE	Kenya	RO	Romania
CA	Canada	KG	Kyrgyzstan	RU	Russian Federation
CF	Central African Republic	KP	Democratic People's Republic of Korea	SD	Sudan
CG	Congo	KR	Republic of Korea	SE	Sweden
CH	Switzerland	KZ	Kazakhstan	SI	Slovenia
CI	Côte d'Ivoire	LI	Liechtenstein	SK	Slovakia
CM	Cameroon	LK	Sri Lanka	SN	Senegal
CN	China	LU	Luxembourg	TD	Chad
CS	Czechoslovakia	LV	Latvia	TG	Togo
CZ	Czech Republic	MC	Monaco	TJ	Tajikistan
DE	Germany	MD	Republic of Moldova	TT	Trinidad and Tobago
DK	Denmark	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	US	United States of America
FI	Finland	MN	Mongolia	UZ	Uzbekistan
FR	France			VN	Viet Nam
GA	Gabon				

Bleaching process comprising use of a phenol oxidizing enzyme, a hydrogen peroxide source and an enhancing agent.

FIELD OF INVENTION

The present invention relates to a process for providing a bleached look in the colour density of the surface of dyed fabric, especially cellulosic fabric such as denim.

BACKGROUND ART

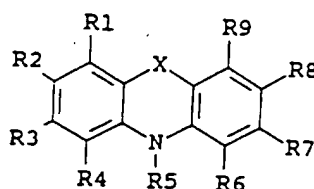
The most usual method of providing a bleached stone-washed look in denim fabric or jeans is by washing the denim or jeans made from such fabric in the presence of pumice stones to provide the desired localized lightening of the colour of the fabric. This is then followed by a bleaching process where the fabric is treated with sodium hypochlorite at 60°C and pH 11-12 for up to 20 min., followed by a neutralisation step and a rinsing. Use of hypochlorite is undesirable, both because chlorite itself is undesirable and because the neutralisation subsequently generates high amounts of salts leading to disposal and pollution problems.

Bleaching enzymes such as peroxidases together with hydrogen peroxide or oxidases together with oxygen have also been suggested for bleaching of dyed textiles (see WO 92/18683), either alone or together with a phenol such as p-hydroxycinnamic acid, 2,4-dichlorophenol, p-hydroxybenzene sulphonate, vanillin or p-hydroxybenzoic acid. The disclosed process is not efficient as can be seen from Example 1 of the present invention.

Thus there is still a need for providing a bleached look in dyed fabrics. The problem to be solved is not easy as many VAT-dyes, especially indigo, are not soluble in water and have a very compact structure on the fibre surface, making them difficult for an enzyme to attack.

SUMMARY OF THE INVENTION

Surprisingly it has been found that it is possible to create a very efficient process for providing a bleached look in the colour density of the surface of dyed fabric, the process comprising contacting, in an aqueous medium, a dyed fabric with a phenol oxidizing enzyme system and an enhancing agent of the following formula:



in which formula X represents (-O-) or (-S-), and the substituent groups R¹-R⁹, which may be identical or different, independently represents any of the following radicals: hydrogen, halogen, hydroxy, formyl, carboxy, and esters and salts hereof, carbamoyl, sulfo, and esters and salts hereof, sulfamoyl, nitro, amino, phenyl, C₁-C₁₄-alkyl, C₁-C₅-alkoxy, carbonyl-C₁-C₅-alkyl, aryl-C₁-C₅-alkyl; which carbamoyl, sulfamoyl, and amino groups may furthermore be unsubstituted or substituted once or twice with a substituent group R¹⁰; and which phenyl may furthermore be unsubstituted or substituted with one or more substituent groups R¹⁰; and which C₁-C₁₄-alkyl, C₁-C₅-alkoxy, carbonyl-C₁-C₅-alkyl, and aryl-C₁-C₅-alkyl groups may be saturated or unsaturated, branched or unbranched, and may furthermore be unsubstituted or substituted with one or more substituent groups R¹⁰;

which substituent group R¹⁰ represents any of the following radicals: halogen, hydroxy, formyl, carboxy and esters and salts hereof, carbamoyl, sulfo and esters and salts hereof, sulfamoyl, nitro, amino, phenyl, aminoalkyl, piperidino, piperazinyl, pyrrolidin-1-yl, C₁-C₅-alkyl, C₁-C₅-alkoxy; which carbamoyl, sulfamoyl, and amino groups may furthermore be unsubstituted or substituted once or twice with hydroxy, C₁-C₅-alkyl, C₁-C₅-alkoxy; and which phenyl may further-

more be substituted with one or more of the following radicals: halogen, hydroxy, amino, formyl, carboxy and esters and salts hereof, carbamoyl, sulfo and esters and salts hereof, and sulfamoyl; and which C₁-C₅-alkyl, and C₁-C₅-alkoxy groups may
5 furthermore be saturated or unsaturated, branched or unbranched, and may furthermore be substituted once or twice with any of the following radicals: halogen, hydroxy, amino, formyl, carboxy and esters and salts hereof, carbamoyl, sulfo and esters and salts hereof, and sulfamoyl;

10 or in which general formula two of the substituent groups R¹-R⁹ may together form a group -B-, in which B represents any of the following the groups: (-CHR¹⁰-N=N-), (-CH=CH-)_n, (-CH=N-)_n or (-N=CR¹⁰-NR¹¹-), in which groups n represents an integer of from 1 to 3, R¹⁰ is a substituent group
15 as defined above and R¹¹ is defined as R¹⁰.

DETAILED DESCRIPTION OF THE INVENTION

Dyed Fabric

The process of the invention is most beneficially applied to cellulose-containing fabrics, such as cotton,
20 viscose, rayon, ramie, linen, Tencel, or mixtures thereof, or mixtures of any of these fibres, or mixtures of any of these fibres together with synthetic fibres such as mixtures of cotton and spandex (stretch-denim). In particular, the fabric is denim. The process of the invention may also be applied to
25 other natural materials such as silk.

The fabric may be dyed with vat dyes such as indigo, or indigo-related dyes such as thioindigo.

In a most preferred embodiment of the process of the invention, the fabric is indigo-dyed denim, including clothing
30 items manufactured therefrom.

Phenol Oxidizing Enzyme Systems

By the term "a phenol oxidizing enzyme system" is meant a system in which an enzyme, by using hydrogen peroxide or molecular oxygen, is capable of oxidizing organic compounds

containing phenolic groups. Examples of such enzymes are peroxidases and oxidases.

If the phenol oxidizing enzyme system requires a source of hydrogen peroxide, the source may be hydrogen peroxide or a hydrogen peroxide precursor for in situ production of hydrogen peroxide, e.g. percarbonate or perborate, or a hydrogen peroxide generating enzyme system, e.g. an oxidase and a substrate for the oxidase, or an amino acid oxidase and a suitable amino acid, or a peroxycarboxylic acid or a salt thereof. Hydrogen peroxide may be added at the beginning of or during the process, e.g. in a concentration corresponding to 0.001-25 mM H_2O_2 .

If the phenol oxidizing enzyme system requires molecular oxygen, molecular oxygen from the atmosphere will usually be present in sufficient quantity.

The enzyme of the phenol oxidizing enzyme systems may be an enzyme exhibiting peroxidase activity or a laccase or a laccase related enzyme as described below.

According to the invention the concentration of the phenol oxidizing enzyme in the aqueous medium where the localized variation in the colour density of the surface of the dyed fabric is taking place, may be 0.001-10000 μg of enzyme protein per g denim, preferably 0.1-1000 μg of enzyme protein per g denim, more preferably 1-100 μg of enzyme protein per g denim.

Peroxidases and Compounds possessing Peroxidase Activity

Compounds possessing peroxidase activity may be any peroxidase enzyme comprised by the enzyme classification (EC 1.11.1.7), or any fragment derived therefrom, exhibiting peroxidase activity, or synthetic or semisynthetic derivatives thereof (e.g. porphyrin ring systems or microperoxidases, cf. e.g. US 4,077,768, EP 537,381, WO 91/05858 and WO 92/16634).

Preferably, the peroxidase employed in the method of the invention is producible by plants (e.g. horseradish or soybean peroxidase) or microorganisms such as fungi or bacteria. Some preferred fungi include strains belonging to the subdivision Deuteromycotina, class Hyphomycetes, e.g. Fusarium,

Humicola, Trichoderma, Myrothecium, Verticillium, Arthromyces, Caldariomyces, Ulocladium, Embellisia, Cladosporium or Dreschlera, in particular Fusarium oxysporum (DSM 2672), Humicola insolens, Trichoderma reesei, Myrothecium verrucana 5 (IFO 6113), Verticillium alboatrum, Verticillium dahliae, Arthromyces ramosus (FERM P-7754), Caldariomyces fumago, Ulocladium chartarum, Embellisia alli or Dreschlera halodes.

Other preferred fungi include strains belonging to the subdivision Basidiomycotina, class Basidiomycetes, e.g. 10 Coprinus, Phanerochaete, Coriolus or Trametes, in particular Coprinus cinereus f. microsporus (IFO 8371), Coprinus macrorrhizus, Phanerochaete chrysosporium (e.g. NA-12) or Trametes (previously called Polyporus), e.g. T. versicolor (e.g. PR4 28-A).

15 Further preferred fungi include strains belonging to the subdivision Zygomycotina, class Mycoraceae, e.g. Rhizopus or Mucor, in particular Mucor hiemalis.

Some preferred bacteria include strains of the order Actinomycetales, e.g. Streptomyces spheroides (ATTC 23965), 20 Streptomyces thermoviolaceus (IFO 12382) or Streptoverticillum verticillium ssp. verticillium.

Other preferred bacteria include Bacillus pumilus (ATCC 12905), Bacillus stearothermophilus, Rhodobacter sphaeroides, Rhodomonas palustri, Streptococcus lactis, 25 Pseudomonas putrefaciens (ATCC 15958) or Pseudomonas fluorescens (NRRL B-11).

Further preferred bacteria include strains belonging to Myxococcus, e.g. M. virescens.

The peroxidase may furthermore be one which is 30 producible by a method comprising cultivating a host cell transformed with a recombinant DNA vector which carries a DNA sequence encoding said peroxidase as well as DNA sequences encoding functions permitting the expression of the DNA sequence encoding the peroxidase, in a culture medium under 35 conditions permitting the expression of the peroxidase and recovering the peroxidase from the culture.

Particularly, a recombinantly produced peroxidase is a peroxidase derived from a Coprinus sp., in particular C.

macrorrhizus or C. cinereus according to WO 92/16634, or a variant thereof, e.g., a variant as described in WO 94/12621.

In the context of this invention, peroxidase acting compounds comprise peroxidase active fragments derived from cytochromes, haemoglobin or peroxidase enzymes, and synthetic or semisynthetic derivatives thereof, e.g. iron porphins, iron porphyrins, and iron phthalocyanine and derivatives thereof.

Determination of peroxidase activity: 1 peroxidase unit (PODU) is the amount of enzyme that catalyzes the conversion of 1 μ mol hydrogen peroxide per minute at the following analytical conditions: 0.88 mM hydrogen peroxide, 1.67 mM 2,2'-azinobis(3-ethylbenzothiazoline-6-sulfonate), 0.1 M phosphate buffer, pH 7.0, incubated at 30°C, photometrically followed at 418 nm.

15 Laccase and Laccase Related Enzymes

In the context of this invention, laccases and laccase related enzymes contemplate any laccase enzyme comprised by the enzyme classification (EC 1.10.3.2), any catechol oxidase enzyme comprised by the enzyme classification (EC 1.10.3.1), any bilirubin oxidase enzyme comprised by the enzyme classification (EC 1.3.3.5) or any monophenol monooxygenase enzyme comprised by the enzyme classification (EC 1.14.99.1).

The laccase enzymes are known from microbial and plant origin. The microbial laccase enzyme may be derived from bacteria or fungi (including filamentous fungi and yeasts) and suitable examples include a laccase derivable from a strain of Aspergillus, Neurospora, e.g., N. crassa, Podospora, Botrytis, Collybia, Fomes, Lentinus, Pleurotus, Trametes, (previously called Polyporus), e.g., T. villosa and T. versicolor, Rhizoctonia, e.g., R. solani, Coprinus, e.g., C. plicatilis and C. cinereus, Psatyrella, Myceliophthora, e.g., M. thermophila, Schytalidium, Phlebia, e.g., P. radita (WO 92/01046), or Coriolus, e.g., C.hirsutus (JP 2-238885).

35 The laccase or the laccase related enzyme may furthermore be one which is producible by a method comprising cultivating a host cell transformed with a recombinant DNA

vector which carries a DNA sequence encoding said laccase as well as DNA sequences encoding functions permitting the expression of the DNA sequence encoding the laccase, in a culture medium under conditions permitting the expression of the laccase enzyme, and recovering the laccase from the culture.

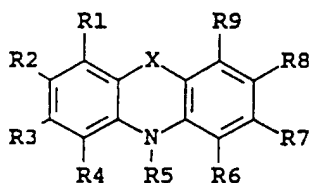
Determination of Laccase Activity (LACU)

Laccase activity is determined from the oxidation of syringaldazin under aerobic conditions. The violet colour produced is photometered at 530 nm. The analytical conditions are 19 μ M syringaldazin, 23.2 mM acetate buffer, pH 5.5, 30°C, 1 min. reaction time.

1 laccase unit (LACU) is the amount of enzyme that catalyses the conversion of 1.0 μ mole syringaldazin per minute at these conditions.

Enhancing Agents

The enhancing agent used in the present invention may be described by the following formula:



in which formula X represents (-O-) or (-S-), and the substituent groups R^1 - R^9 , which may be identical or different, independently represents any of the following radicals: hydrogen, halogen, hydroxy, formyl, carboxy, and esters and salts hereof, carbamoyl, sulfo, and esters and salts hereof, sulfamoyl, nitro, amino, phenyl, C_1 - C_{14} -alkyl, C_1 - C_5 -alkoxy, carbonyl- C_1 - C_5 -alkyl, aryl- C_1 - C_5 -alkyl; which carbamoyl, sulfamoyl, and amino groups may furthermore be unsubstituted or substituted once or twice with a substituent group R^{10} ; and which phenyl may furthermore be unsubstituted or substituted

with one or more substituent groups R^{10} ; and which C_1-C_{14} -alkyl, C_1-C_5 -alkoxy, carbonyl- C_1-C_5 -alkyl, and aryl- C_1-C_5 -alkyl groups may be saturated or unsaturated, branched or unbranched, and may furthermore be unsubstituted or substituted with one or
5 more substituent groups R^{10} ;

which substituent group R^{10} represents any of the following radicals: halogen, hydroxy, formyl, carboxy and esters and salts hereof, carbamoyl, sulfo and esters and salts hereof, sulfamoyl, nitro, amino, phenyl, aminoalkyl,
10 piperidino, piperaziny, pyrrolidin-1-yl, C_1-C_5 -alkyl, C_1-C_5 -alkoxy; which carbamoyl, sulfamoyl, and amino groups may furthermore be unsubstituted or substituted once or twice with hydroxy, C_1-C_5 -alkyl, C_1-C_5 -alkoxy; and which phenyl may furthermore be substituted with one or more of the following radicals:
15 halogen, hydroxy, amino, formyl, carboxy and esters and salts hereof, carbamoyl, sulfo and esters and salts hereof, and sulfamoyl; and which C_1-C_5 -alkyl, and C_1-C_5 -alkoxy groups may furthermore be saturated or unsaturated, branched or unbranched, and may furthermore be substituted once or twice
20 with any of the following radicals: halogen, hydroxy, amino, formyl, carboxy and esters and salts hereof, carbamoyl, sulfo and esters and salts hereof, and sulfamoyl;

or in which general formula two of the substituent groups R^1-R^9 may together form a group -B-, in which B
25 represents any of the following the groups: $(-CHR^{10}-N=N-)$, $(-CH=CH-)_n$, $(-CH=N-)_n$ or $(-N=CR^{10}-NR^{11}-)$, in which groups n-represents an integer of from 1 to 3, R^{10} is a substituent group as defined above and R^{11} is defined as R^{10} . (It is to be understood that if the above mentioned formula comprises two or
30 more R^{10} -substituent groups, these R^{10} -substituent groups may be the same or different).

In particular embodiments, the enhancing agent is 10-methylphenothiazine, phenothiazine-10-propionic acid, N-hydroxysuccinimide phenothiazine-10-propionate, 10-ethyl-
35 phenothiazine-4-carboxylic acid, 10-ethylphenothiazine, 10-propylphenothiazine, 10-isopropylphenothiazine, methyl phenothiazine-10-propionate, 10-phenylphenothiazine, 10-allylphenothiazine, 10-(3-(4-methylpiperazin-1-yl)propyl)phenothiazine,

10-(2-pyrrolidin-1-yl-ethyl)phenothiazine, 2-methoxy-10-methylphenothiazine, 1-methoxy-10-methylphenothiazine, 3-methoxy-10-methylphenothiazine, 3,10-dimethylphenothiazine, 3,7,10-trimethylphenothiazine, 10-(2-hydroxyethyl)phenothiazine, 10-
5 (3-hydroxypropyl)phenothiazine, 3-(2-hydroxyethyl)-10-methylphenothiazine, 3-hydroxymethyl-10-methylphenothiazine, 3,7-dibromophenothiazine-10-propionic acid, phenothiazine-10-propionamide, chlorpromazine, 2-chloro-10-methylphenothiazine, 2-acetyl-10-methylphenothiazine, 10-methylphenoxazine, 10-
10 ethylphenoxazine, phenoxazine-10-propionic acid, 10-(2-hydroxyethyl)phenoxazine or 4-carboxyphenoxazine-10-propionic acid.

The enhancing agent of the invention may be present in concentrations of from 0.005 to 1000 μ mole per g denim, preferably 0.05 to 500 μ mole per g denim, more preferably 0.5
15 to 100 μ mole per g denim.

Stability of the Radical of the Enhancing Agent

Without being limited to any theory it is presently contemplated that there is a positive correlation between the
20 half-life of the radical which the enhancing agent forms in the relevant aqueous medium and its efficiency in providing a bleached look in the colour density of the surface of the dyed fabric together with the phenol-oxidizing enzyme system, and that this half-life is significantly longer than the half-life
25 of any of the substances selected from the group consisting of p-hydroxycinnamic acid, 2,4-dichlorophenol, p-hydroxybenzene sulphonate, vanillin and p-hydroxybenzoic acid (i.e. the enhancing agents disclosed in WO 92/18683).

This invention therefore further relates to a process
30 for providing a bleached look in the colour density of the surface of dyed fabric, the process comprising contacting, in an aqueous medium, a dyed fabric with a phenol oxidizing enzyme system and an enhancing agent, wherein said enhancing agent is capable of forming a radical having a half-life, in said
35 aqueous medium, which is at least 10 times longer than the radical half-life of any one of the substances selected from

the group consisting of p-hydroxycinnamic acid, 2,4-dichlorophenol, p-hydroxybenzene sulphonate, vanillin and p-hydroxybenzoic acid, tested in the same aqueous medium, in particular wherein said enhancing agent is capable of forming
5 a radical having a half-life, in said aqueous medium, which is at least 100 times longer than the radical half-life of any one of the substances selected from the group consisting of p-hydroxycinnamic acid, 2,4-dichlorophenol, p-hydroxybenzene sulphonate, vanillin and p-hydroxybenzoic acid, tested in the
10 same aqueous medium.

As the half-life of the radical is dependent on, inter alia, the pH, the temperature and the buffer of the aqueous medium, it is very important that all these factors are the same when the half-lives of the radicals of various
15 enhancing agents are compared.

Industrial Applications

The process of the present invention is typically used in industrial machines for making fabric look bleached. Normally, the process of the invention will be performed on
20 fabric already stonewashed, but the process may also be applied to fabric which has not undergone a stonewashing process beforehand. Most commonly the fabric is added to the machine according to the machine capacity per the manufacturer's instructions. The fabric may be added to the machine prior to
25 introducing water or the fabric may be added after water is introduced. The phenol oxidizing enzyme system and the enhancing agent of the invention may be present in the water prior to adding the fabric or they may be added after the fabric has been wetted. The phenol oxidizing enzyme system may
30 be added simultaneously with the enhancing agent or they may be added separately. After the fabric has been contacted with the phenol oxidizing enzyme system and the enhancing agent of the invention it should be agitated in the machine for a sufficient period of time to ensure that the fabric is fully wetted and to
35 ensure the action of the enzyme system and the enhancing agent.

We have found (see the Examples below) that the optimum bleaching conditions might be a compromise between

optimum stability of the enzyme, optimum activity of the enzyme, optimum stability of the radical of the enhancing agent, and optimum reactivity (oxidation potential) of the radical, as well as choice of buffering system (buffer capacity, buffer toxicity, costs of buffer etc.).

The invention is further illustrated in the following examples which are not intended to be in any way limiting to the scope of the invention as claimed.

EXAMPLE 1

10 Bleaching Denim with Laccase and Different Enhancing Agents

The test procedure for denim bleaching was performed as described below:

Enhancing agents: The enhancing agents were obtained from Sigma-Aldrich, Janssen Chimica, Kodak, Tokyo Kasai Organic
15 Chemicals, Daiichi Pure Chemicals Co. or Boehringer Mannheim; N-methylated derivatives of phenothiazine and phenoxazine may be prepared by methylation with methyl iodide as described by Cornel Bodea and Ioan Silberg in "Recent Advances in the Chemistry of Phenothiazines" (Advances in heterocyclic
20 chemistry, 1968, Vol. 9, pp. 321-460); B. Cardillo & G. Casnati in Tetrahedron, 1967, Vol. 23, p. 3771. Phenothiazine and phenoxazine propionic acids may be prepared as described in J. Org. Chem. 15, 1950, pp. 1125-1130. Hydroxyethyl and hydroxypropyl derivatives of phenothiazine and phenoxazine may
25 be prepared as described by G. Cauquil in Bulletin de la Society Chimique de France, 1960, p.1049.

Enzyme: Laccase derived from Trametes villosa (SP 504, available from Novo Nordisk A/S) was used.

Procedure: 18 ml 0.01 M B&R (Britt & Robinson) buffer (pH 4, 6,
30 or 8) were added to a 50 ml conical flask. A magnet bar (4 cm) and a circular piece of stone washed denim (3.5 cm diameter

0.4 g) were added to the flask together with 1 ml of the stock solution of the enhancing agent to be tested and 1 ml of enzyme, giving a denim:liquor (w/w) ratio of 1:50; the final concentrations of the enhancing agent and the enzyme shown in Table 1-2 below.

The flask was incubated for 2-3 hours on a magnet stirrer in a water bath (50°C and approximately 200 rpm). After the enzymatic bleaching, the denim swatch was rinsed with distilled water and air dried, whereafter it was evaluated for the degree of bleaching. The evaluation was performed visually and by using a Minolta Chroma Meter CR200 or a Minolta Chroma Meter CR300.

Evaluation: A Minolta Chroma Meter CR200 or CR300 (available from Minolta Corp.) was used according to Manufacturer's instructions to evaluate the degree of bleaching as well as to estimate any discoloration using the change in the colour space coordinates $L^*a^*b^*$ (CIELAB-system): L^* gives the change in white/black at a scale of from 0 to 100, a^* gives the change in green ($-a^*$)/red ($+a^*$), and b^* gives the change in blue ($-b^*$)/yellow ($+b^*$). A decrease in L^* means an increase in black colour (decrease of white colour), an increase in L^* means an increase in white colour (a decrease in black colour), a decrease in a^* means an increase in green colour (decrease in red colour), an increase in a^* means an increase in red colour (a decrease in green colour), a decrease in b^* means an increase in blue colour (a decrease in yellow colour), and an increase in b^* means an increase in yellow colour (a decrease in blue colour).

The bleached stone washed denim swatches were compared to non-treated stone washed denim swatches.

The Minolta Chroma Meter CR200 or the Minolta Chroma Meter CR300 was operated in the $L^*a^*b^*$ colour space (coordinate system). The light source used was a CIE light standard C. Each measurement was an average of 3 measurements. The instrument was calibrated using a Minolta calibration plate (white). 10 non-treated denim swatches were measured 2 times each and the average of the coordinates $L^*a^*b^*$ were calculated and entered as

a reference. The coordinates of the samples were then calculated as the difference (Δ) of the average of 3 measurements on each swatch from the reference value of the coordinates $L^*a^*b^*$.

5 Table 1

Table 1 shows $\Delta(L^*/a^*/b^*)$ between a swatch treated with the tested system and a non-treated swatch at pH 4, 6 and 8.

Tested System	pH 4	pH 6	pH 8
10 Phenoxazine-10-propionic acid (3 hours): (1000 μ M - 50 μ mole/g) (1.0 LACU/ml - 15 780 μ g/g) (2 hours): (100 μ M - 5 μ mole/g) (0.1 LACU/ml - 20 78 μ g/g)	25.8/2.6/33.7	32.6/2.6/33.1 5.5/-1.0/1.9	6.4/-1.8/2.4
25 Phenoxazine-10-hydroxyethyl (3 hours): (1000 μ M - 50 μ mole/g) (1.0 LACU/ml - 780 μ g/g)	23.9/6.5/33.6	18.9/-0.1/- 29.2	3.3/-0.8/1.6
30 Phenothiazine-10-ethyl-4-carboxy (3 hours): (1000 μ M - 50 μ mole/g) (1.0 LACU/ml - 35 780 μ g/g)	11.9/-1.7/2.8	20.6/-2.9/5.8	2.0/-0.3/0.5
40 Phenothiazine-10-propionic acid (3 hours): (1000 μ M - 50 μ mole/g) (1.0 LACU/ml - 780 μ g/g)	14.9/-2.3/3.7	11.6/-1.8/3.0	5.6/-1.1/0.8

5 Promazine hydrochloride (3 hours): (1000 μ M - 50 μ mole/g) (0.1 LACU/ml - 78 μ g/g)	16.1/-1.8/4.6	8.1/-1.2/3.3	-2.3/0.7/0.0
10 Phenothiazine- 10- ethylalcohol (3 hours): (1000 μ M - 50 μ mole/g) (1.0 LACU/ml - 15 780 μ g/g)	19.7/-2.4/4.9	15.7/-1.9/4.2	4.6/-0.6/0.5

Visually a ΔL^* around 5 gives a significant effect so it can be seen from the results presented in Table 1 that all the tested systems have a significant effect at pH 4-6 in bleaching denim.

20 Table 2

Table 2 shows $\Delta(L^*/a^*/b^*)$ between a swatch treated with the enhancing agents described in WO 92/18683 + laccase (0.1-1.0 LACU/ml corresponding to 78 μ g enzyme protein/g denim - 780 μ g enzyme protein/g denim) and a non-treated swatch at pH 4, 6
25 and 8.

	Tested System	pH 4	pH 6	pH 8
5	p-Hydroxy-benzoic acid: (1000 μ M - 50 μ mole/g) Laccase: (0.1 LACU/ml - 78 μ g/g)	0.85/ -0.09/ 0.61	0.91/ -0.19/ -0.14	-0.21/ 0.24/ -0.17
10	p-Hydroxy-benzene-sulfonate: (1000 μ M - 50 μ mole/g) Laccase: (0.1 LACU/ml - 78 μ g/g)	-0.18/ 0.14/ -0.12	0.33/ 0.06/ -0.22	-0.51/ 0.17/ -0.20
20	2,4-Dichloro-phenol: (1000 μ M - 50 μ mole/g) Laccase: (0.1 LACU/ml - 78 μ g/g)	0.64/ -0.22/ 0.5	-0.19/ -0.19/ 0.57	-0.54/ 0.16/ -0.14
25	Vanillin: (1000 μ M - 50 μ mole/g) Laccase: (1.0 LACU/ml - 780 μ g/g)	-0.67/ -0.34/ 1.41	0.28/ -0.03/ 0.49	-0.38/ -0.05/ 0.75
30	p-Hydroxy-cinnamic acid: (1000 μ M - 50 μ mole/g) Laccase: (1.0 LACU/ml - 780 μ g/g)	0.64/ -0.53/ 1.62	4.47/ -0.63/ 3.88	2.97/ -0.45/ 0.79

From the results presented in Table 2 it can be seen that none of the prior art described enhancing agents have any significant effect in bleaching the denim.

EXAMPLE 2Bleaching Denim with Laccase and Phenothiazine-10-Propionic Acid using Different Buffers

To illustrate the effect of different buffers on the
5 denim bleaching performance the following tests have been made:

11 different buffers and 3 types of water were tested. Each buffer was prepared at a concentration of 0.01 M, and pH adjusted to pH 6.5 with NaOH or with the corresponding acid. 80 ml of the buffer in question was added to a 200 ml
10 glass beaker together with a magnet bar (4 cm), and 8 circular pieces of denim (3.5 cm in diameter ~ 0.4 g), giving a denim:liquor ratio of 1:25.

The glass beaker was incubated on a magnet stirrer (300 rpm) in a water bath at 60°C, and a pH electrode was
15 dipped into the liquor in the middle of the beaker in order to monitor and control pH at pH 6.5 (i.e. the experiments were run under pH-stat conditions using a Radiometer pH-stat (PHM 82 or PHM 62 pH meter, TTT 80 Titrator, ABU 80 Autoburette) with automatic titration with the corresponding acid (0.1 M) if and
20 when pH increased above pH 6.5). Following equilibration at pH 6.5, phenothiazine-10-propionic acid (PPT), 0.02 M in 96% ethanol, was added to a final concentration of 250 μ M ~ 6.3 μ mole/g together with laccase from Trametes villosa (TVL) (20 LACU/ml in water, available from Novo Nordisk A/S) to a
25 final concentration of 0.1 LACU/ml ~ 39 μ g/g. After 30 minutes the denim swatches were rinsed in tap-water and air dried on filter paper overnight, and the resulting degree of bleaching was determined as mentioned above in Example 1. The results are shown in Table 3.

Table 3

Bleaching obtained using 250 μ M PPT \sim 6.3 μ mole/g and 0.1 LACU/ml \sim 39 μ g/g of TvL at pH 6.5 (pH-stat) in 30 minutes at 60°C in different buffer systems, all 0.01 M (except the systems using various sources of water). pH was continuously monitored and controlled at pH 6.5 titrating with the corresponding acid, except that for borate buffer and glycine buffer pH was controlled titrating with 0.1 M HCl, and for de-ionized water, for Milli Q UF water, and for tap water, pH was controlled titrating with 0.1 M H₂SO₄.

Buffer	Degree of bleaching (ΔL^*)
Oxalate	13.12
Borate	11.10
De-ionized water	10.38
15 Acetate	10.17
Glycine	10.05
Milli Q UF water	10.04
Cold tap water	9.26
Maleic	8.39
20 Succinic	7.34
3,3-Dimethylglutaric	6.69
B&R	6.55
Phosphate	6.44
Citrate/phosphate	6.35
25 Citrate	3.15

The results obtained in Table 3 are in accordance with results obtained determining PPT radical stability ($T_{1/2}$) in various buffers at various pH following the general correlation: A high radical stability will give a high bleaching performance, and a low radical stability will give a low bleaching performance.

EXAMPLE 3

Bleaching Denim with Laccase and Phenothiazine-10-Propionic Acid at different pH values

To illustrate pH's influence on the denim bleaching process, a pH profile was made in the following way:

0.01 M oxalate buffer was adjusted to the appropriate pH in the range pH 4.0 - pH 7.5 using oxalic acid or oxalate. 80 ml buffer was added to a 200 ml glass beaker together with a magnet bar (4 cm), and 8 circular pieces of denim (3.5 cm in diameter \times 0.4 g), giving a denim:liquor ratio of 1:25. The glass beaker was incubated on a magnet stirrer (300 rpm) in a water bath at 50°C, and a pH electrode was dipped into the liquor in the middle of the beaker in order to monitor and control pH at the desired pH in the range 4.0-7.5 (i.e. the experiments were run under pH-stat conditions using a Radiometer pH-stat (PHM 82 or PHM 62 pH meter, TTT 80 Titrator, ABU 80 Autoburette) with automatic titration with 0.1 M oxalic acid if and when pH increased above set-point).

Following equilibration at the desired pH, phenothiazine-10-propionic acid (PPT), 0.02 M in 96% ethanol, was added to a final concentration of 83.3 μ M \sim 2.1 μ mole/g together with laccase from Trametes villosa (TvL) or Myceliophthora thermophila (MtL); TvL available from Novo Nordisk A/S and MtL produced as described in PCT/US95/06815, to a final concentration of 0.1 LACU/ml \sim 39 μ g/g (TvL) and 54 μ g/g (MtL).

After 10 and 20 minutes PPT was added corresponding to 83.3 μ M \sim 2.1 μ mole/g (the total amount of PPT used is 250 μ M \sim 6.3 μ mole/g).

After 30 minutes the denim swatches were rinsed in tap-water and air dried on filter paper overnight, and the resulting degree of bleaching was determined as mentioned above in Example 1. The results are shown in Table 4.

5 Table 4

Bleaching obtained using 250 μ M PPT ($3 \times 83.3 \mu$ M) \sim 6.3 μ mole/g and 0.1 LACU/ml of TvL or MtL \sim 39 μ g/g (TvL) and 54 μ g/g (MtL), at pH 4.0-7.5 (pH-stat) in 30 minutes at 50°C in 0.01 M oxalate buffer. pH was continuously monitored and
 10 controlled at set-point pH titrating with 0.1 M oxalic acid.

pH	Degree of bleaching (ΔL^*)	
	TvL	MtL
4.0	0.98	1.27
4.5	3.16	2.62
5.0	2.77	4.70
15 5.5	4.36	6.88
6.0	5.65	5.45
6.5	6.79	4.94
7.0	6.36	1.76
7.5	2.82	1.00

20 It can be seen from Table 4 that when using the above described conditions the pH-optimum of the denim bleaching process for the T. villosa laccase is around pH 6.5 and the pH-optimum of the denim bleaching process for the M. thermophila laccase is around pH 5.5.

EXAMPLE 4Bleaching Denim with Laccase and Phenothiazine-10-Propionic Acid at Different Temperatures

To illustrate the influence of temperature on the denim bleaching process, a temperature profile was made in the following way:

0.01 M oxalate buffer was adjusted to the appropriate pH using oxalic acid or oxalate. 80 ml buffer was added to a 200 ml glass beaker together with a magnet bar (4 cm), and 8 circular pieces of denim (3.5 cm in diameter ~ 0.4 g), giving a denim:liquor ratio of 1:25. The glass beaker was incubated on a magnet stirrer (300 rpm) in a water bath at the appropriate temperature in the range 30°C-80°C, and a pH electrode was dipped into the liquor in the middle of the beaker in order to monitor and control pH at the desired pH (i.e. the experiments were run under pH-stat conditions using a Radiometer pH-stat (PHM 82 or PHM 62 pH meter, TTT 80 Titrator, ABU 80 Autoburette) with automatic titration with 0.1 M oxalic acid if and when pH increased above set-point). Following equilibration at the desired pH, phenothiazine-10-propionic acid (PPT), 0.02 M in 96% ethanol, was added to a final concentration of 83.3 μ M ~ 2.1 μ mole/g together with laccase from Trametes villosa (TvL) or Myceliophthora thermophila (MtL); TvL available from Novo Nordisk A/S and MtL produced as described in PCT/US95/06815, to a final concentration of 0.1 LACU/ml ~ 39 μ g/g (TvL) and 54 μ g/g (MtL).

After 10 and 20 minutes PPT was added corresponding to 83.3 μ M ~ 2.1 μ mole/g (the total amount of PPT is 6.3 μ mole/g). After 30 minutes the denim swatches were rinsed in tap-water and air dried on filter paper overnight, and the resulting degree of bleaching was determined as mentioned above in Example 1. The results are shown in Table 5.

Table 5

Bleaching obtained using 250 μ M PPT (3x83.3 μ M) ~ 6.3 μ mole/g and 0.1 LACU/ml of TvL or MtL ~ 39 μ g/g (TvL) or 54 μ g/g, at pH 6.5 and pH 5.5 respectively in 30 minutes in 0.01 M oxalate buffer. pH was continuously monitored and controlled at set-point pH titrating with 0.1 M oxalic acid.

Temperature °C	Degree of bleaching (ΔL^*)	
	TvL (pH 6.5)	MtL (pH 5.5)
30	3.27	4.29
40	6.12	5.36
50	6.59	6.76
60	7.58	7.68
70	5.92	7.80
80	2.68	4.48

It can be seen from Table 5 that when using the above described conditions the temperature-optimum of the denim bleaching process for the T. villosa laccase is around 60°C and the temperature-optimum of the denim bleaching process for the M. thermophila laccase is around 60-70°C.

EXAMPLE 5

Enzyme Dosage Response in the Denim Bleaching Process

To illustrate the enzyme dosage response in the denim bleaching process, an enzyme dosage response profile was made in the following way:

0.01 M oxalate buffer was adjusted to the appropriate pH using oxalic acid or oxalate. 80 ml buffer was added to a

200 ml glass beaker together with a magnet bar (4 cm), and 8 circular pieces of denim (3.5 cm in diameter \times 0.4 g), giving a denim:liquor ratio of 1:25. The glass beaker was incubated on a magnet stirrer (300 rpm) in a water bath at the appropriate temperature, and a pH electrode was dipped into the liquor in the middle of the beaker in order to monitor and control pH at the desired pH (i.e. the experiments were run under pH-stat conditions using a Radiometer pH-stat (PHM 82 or PHM 62 pH meter, TTT 80 Titrator, ABU 80 Autoburette) with automatic titration with 0.1 M oxalic acid if and when pH increased above set-point). Following equilibration at the desired pH, phenothiazine-10-propionic acid (PPT), 0.02 M in 96% ethanol, was added to a final concentration of $83.3 \mu\text{M}$ \times 2.1 $\mu\text{mole/g}$ together with laccase from Trametes villosa (TvL) or Myceliophthora thermophila (MtL); TvL available from Novo Nordisk A/S and MtL produced as described in PCT/US95/06815.

After 10 and 20 minutes PPT was added corresponding to $83.3 \mu\text{M}$ \times 2.1 $\mu\text{mole/g}$, giving a total amount of PPT of 6.3 $\mu\text{mole/g}$. After 30 minutes the denim swatches were rinsed in tap-water and air dried on filter paper overnight, and the resulting degree of bleaching was determined as mentioned above in Example 1. The results are shown in Table 6.

Table 6

Bleaching obtained using 250 μM PPT ($3 \times 83.3 \mu\text{M}$) \times 6.3 $\mu\text{mole/g}$ and different concentrations of TvL or MtL at pH 6.5 and pH 5.5, respectively, and at 60°C and 70°C, respectively in 30 minutes in 0.01 M oxalate buffer. pH was continuously monitored and controlled at set-point pH titrating with 0.1 M oxalic acid.

Enzyme Dosage	Degree of bleaching (ΔL^*)	
	TvL (pH 6.5, 60°C)	MtL (pH 5.5, 70°C)
0.01 LACU/ml ~ 3.9 $\mu\text{g/g}$ (TvL) or 5.4 $\mu\text{g/g}$ (MtL)	3.70	5.19
0.05 LACU/ml ~ 19.5 $\mu\text{g/g}$ (TvL) or 27 $\mu\text{g/g}$ (MtL)	8.93	8.30
0.1 LACU/ml ~ 39 $\mu\text{g/g}$ (TvL) or 54 $\mu\text{g/g}$ (MtL)	10.97	9.43
0.5 LACU/ml ~ 195 $\mu\text{g/g}$ (TvL) or 272 $\mu\text{g/g}$ (MtL)	14.32	8.91
1.0 LACU/ml ~ 390 $\mu\text{g/g}$ (TvL) or 540 $\mu\text{g/g}$ (MtL)	12.98	6.96

It can be seen from Table 6 that when using the above described conditions both enzymes exhibit a typical enzyme dosage response profile, and that the enzyme dosage optimum of the denim bleaching process for the *T. villosa* laccase is around 0.5 LACU/ml ~ 195 $\mu\text{g/g}$ and the enzyme dosage optimum of the denim bleaching process for the *M. thermophila* laccase is around 0.1 LACU/ml ~ 54 $\mu\text{g/g}$.

EXAMPLE 6

The Bleach Response as a Function of Time in the Denim Bleaching Process

To illustrate the bleach response as a function of time in the denim bleaching process, a time profile was made in the following way:

Two different buffers were used (B&R buffer and oxalate buffer). Each buffer was prepared at a concentration of 0.01 M, and pH adjusted to the appropriate pH with NaOH or with the corresponding acid. 20 ml of the buffer in question was

added to a 50 ml conical flask together with a magnet bar (4 cm), and 2 circular pieces of denim (3.5 cm in diameter - 0.4 g), giving a denim:liquor ratio of 1:25. The flasks were incubated on a magnet stirrer (300 rpm) in a water bath at 60°C. Following equilibration, phenothiazine-10-propionic acid (PPT) was added to a final concentration of 250 μ M (0.02 M in 96% ethanol) - 6.3 μ mole/g together with laccase from Trametes villosa (TVL) to a final concentration of 0.1 LACU/ml - 39 μ g/g, TVL available from Novo Nordisk A/S. Following bleaching the denim swatches were rinsed in tap water and air dried on filter paper overnight, and the resulting degree of bleaching was determined as mentioned above in Example 1.

6 identical flasks were made with each buffer system, and the degree of bleaching was determined after 5, 10, 15, 30, 45, and 60 minutes, respectively. The results are shown in Table 7.

Table 7

Bleaching obtained as a function of time for different buffer systems using 250 μ M PPT - 6.3 μ mole/g (added at the beginning of the experiment), 0.1 LACU/ml - 39 μ g/g of TVL, at 60°C.

Time (minutes)	Degree of bleaching (ΔL^*)	
	0.01 M B&R Buffer Initial pH: 6.0 Final pH: 6.2	0.01 M Oxalate buffer Initial pH: 5.0 Final pH: 6.7-7.0
(0)	(0)	(0)
5	4.48	5.49
10	7.66	7.58
15	8.18	8.28
30	8.90	10.50
45	9.21	12.73
60	9.27	11.46

It can be seen from Table 7 that when using the above described conditions the bleaching process proceeds very fast for the first 10-15 minutes, and that the optimum bleaching time of the denim bleaching process for the T. villosa laccase is around 60 min. and around 45 min. when using B&R buffer and oxalate buffer, respectively.

EXAMPLE 7

Denim Bleaching in Larger Scale (300 ml) using $(\text{NH}_4)_2\text{SO}_4/\text{NaHSO}_4$ as buffer

10 Experiments were performed in larger scale (300 ml scale) in a launder-ometer. A pH profile in 20 mM $(\text{NH}_4)_2\text{SO}_4/\text{NaHSO}_4$ was made.

An Atlas LP2 launder-ometer was used. 300 ml 0.02 M $(\text{NH}_4)_2\text{SO}_4/\text{NaHSO}_4$ was adjusted to the appropriate pH in the range
15 pH 1.5-7.0. 300 ml buffer was added to a 1200 ml beaker together with 12 g denim (in one piece), giving a denim:liquor ratio of 1:25; additionally 30 LACU - 469 μg Trametes villosa laccase (TVL - available from Novo Nordisk A/S) and 0.020 g phenothiazine-10-propionic acid (PPT) were added resulting in
20 a laccase concentration of 39 $\mu\text{g/g}$ and a PPT concentration of 6.2 $\mu\text{mole/g}$.

The beakers were sealed and placed in the launder-ometer and processed for 55 minutes (15 minutes heating time 22°C-60°C, 40 minutes holding time). After processing, samples
25 of the processing liquor were diluted in methanol (10-25 x) and analyzed for residual amount of PPT by HPLC.

The HPLC method was based on the following: Column: Supelcosil LC-18-DB, RP C-18, 3.6x250 mm, Eluent: 70% methanol, 30% 25 mM PO_4 buffer pH 6.5, Flow: 1.0 ml/min, Detection: UV/Vis
30 diode array (monitoring at 238, 296, and 600 nm), Injection: 20 μl , Sample dilution: Methanol.

The results obtained are shown in Table 8.

Table 8

Bleaching obtained in launder-ometer as a function of pH. Conditions: 300 ml 0.02 M $(\text{NH}_4)_2\text{SO}_4/\text{NaHSO}_4$ buffer was added to a 1200 ml beaker together with 12 g denim (in one piece), 30 LACU 5 $\sim 469 \mu\text{g}$ TvL and 0.020 g PPT. The beakers were sealed and placed in the launder-ometer and processed for 55 minutes (15 minutes heating time 22°C-60°C, 40 minutes holding time).

Initial pH	Final pH	ΔL^*	Residual PPT (μM)
1.56	1.57	2.66	58 \sim 1.5 $\mu\text{mole/g}$
2.02	2.08	2.02	77 \sim 1.9 $\mu\text{mole/g}$
2.48	2.67	2.82	48 \sim 1.2 $\mu\text{mole/g}$
2.78	3.18	4.59	0
2.99	3.70	9.51	29 \sim 0.7 $\mu\text{mole/g}$
3.15	4.34	13.10	0
3.28	5.22	15.33	0
3.41	5.75	14.93	0
3.61	6.00	18.36	0
3.81	6.15	13.81	0
4.03	6.28	17.31	23 \sim 0.6 $\mu\text{mole/g}$
5.06	6.57	17.20	43 \sim 1.1 $\mu\text{mole/g}$
5.97	7.24	9.04	200 \sim 5.0 $\mu\text{mole/g}$
6.13	7.36	8.67	212 \sim 5.3 $\mu\text{mole/g}$
6.70	7.89	4.31	273 \sim 6.9 $\mu\text{mole/g}$
7.03	8.02	3.30	284 \sim 7.2 $\mu\text{mole/g}$

It can be seen from Table 8 that a high degree of bleaching is achieved at an initial pH in the range of from 3.3 to 5.1, the highest degree of bleaching being at an initial pH value of about 3.6.

5 EXAMPLE 8

Denim Bleaching in Larger Scale (300 ml) using acetate as buffer

Experiments similar to the series using $(\text{NH}_4)_2\text{SO}_4/\text{NaHSO}_4$ (see Example 7) were made in acetate buffer.
10 Conditions: An Atlas LP2 launder-ometer was used. 300 ml 10 mM acetate buffer pH 3.5-6.5 was added to a 1200 ml beaker together with 12 g denim (in one piece), giving a denim:liquor ratio of 1:25. Additionally 30 LACU - 469 μg Trametes villosa laccase (TvL) or 30 LACU - 652 μg Myceliophthora thermophila
15 laccase (MtL) - TvL available from Novo Nordisk A/S and MtL produced as described in PCT/US95/06815 - and 0.02 g phenothiazine-10-propionic acid (PPT) were added. The conditions were thus: 39 $\mu\text{g/g}$ TvL or 54 $\mu\text{g/g}$ MtL and 6.3 $\mu\text{mole/g}$ PPT.

20 The beakers were sealed and placed in the launder-ometer and processed for 40 minutes (10 minutes heating time 22°C-60°C, 30 minutes holding time). After processing, samples of the processing liquor were diluted in methanol (10-25 x) and analyzed for residual amount of PPT by HPLC.

25 The HPLC method was based on the following: Column: Supelcosil LC-18-DB, RP C-18, 3.6x250 mm, Eluent: 70% methanol, 30% 25 mM PO_4 buffer pH 6.5, Flow: 1.0 ml/min, Detection: UV/Vis diode array (monitoring at 238, 296, and 600 nm), Injection: 20 μl , Sample dilution: Methanol. The results obtained are shown
30 in Tables 9-10 below.

Table 9

Bleaching obtained in launder-ometer as a function of pH. Conditions: 300 ml 0.01 M acetate buffer was added to a 1200 ml beaker together with 12 g denim (in one piece), 30 LACU TvL (to
35 0.1 LACU/ml), and 0.020 g PPT (to 250 μM). The beakers were

sealed and placed in the launder-ometer and processed for 40 minutes (10 minutes heating time 22°C-60°C, 30 minutes holding time).

Initial pH	Final pH	ΔL^*	Residual PPT (μM)
3.50	3.58	4.49	5 - 0.1 $\mu mole/g$
4.00	4.07	7.21	16 - 0.4 $\mu mole/g$
4.50	4.58	7.77	10 - 0.3 $\mu mole/g$
5.00	5.10	11.29	0
5.50	5.91	13.51	16 - 0.4 $\mu mole/g$
6.00	7.10	3.2	239 - 6.0 $\mu mole/g$
6.50	7.61	2.47	242 - 6.1 $\mu mole/g$
7.67 ¹⁾	8.23 ¹⁾	2.35 ¹⁾	258 ¹⁾ - 6.5 $\mu mole/g$

¹⁾: No buffer added, only tap water, PPT and laccase.

It can be seen from Table 9 that a very high degree of bleaching is achieved at an initial pH value of about 5.5.

Table 10

Bleaching obtained in launder-ometer as a function of pH. Conditions: 300 ml 10 mM acetate buffer was added to a 1200 ml beaker together with 12 g denim (in one piece), 30 LACU MtL (to 20 0.1 LACU/ml), and 0.020 g PPT (to 250 μM). The beakers were sealed and placed in the launder-ometer and processed for 40 minutes (10 minutes heating time 22°C-60°C, 30 minutes holding time).

Initial pH	Final pH	ΔL^*	Residual PPT (μM)
3.50	3.58	5.02	33 - 0.8 $\mu mole/g$
4.00	4.06	7.87	12 - 0.3 $\mu mole/g$
4.50	4.57	9.31	8 - 0.2 $\mu mole/g$
5.00	5.12	12.07	12 - 0.3 $\mu mole/g$
5.50	5.96	15.16	49 - 1.2 $\mu mole/g$
6.00	7.43	8.65	204 - 5.1 $\mu mole/g$
6.50	7.92	7.8	237 - 6.0 $\mu mole/g$

It can be seen from Table 10 that a very high degree
10 of bleaching is achieved at an initial pH value of about 5.5.

EXAMPLE 9

Dosage Response with Respect to Phenothiazine-10-Propionic Acid (PPT) in Larger Scale (300 ml)

To illustrate the dosage-response with respect to
15 PPT, a series of experiments in launder-ometer scale was
performed. An Atlas LP2 launder-ometer was used. 300 ml 20 mM
(NH_4)₂SO₄/NaHSO₄ pH 5.4 was added to a 1200 ml beaker together
with 12 g denim (in one piece), 30 LACU *Trametes villosa* (TVL)
- available from Novo Nordisk A/S - (to 0.1 LACU/ml), and PPT

in the range of from 50 μM to 500 μM . The conditions were thus: a denim:liquor ratio of 1:25; 39 $\mu\text{g/g}$ denim TvL and 1.3 $\mu\text{mole/g}$ denim - 12.5 $\mu\text{mole/g}$ denim PPT.

The beakers were sealed and placed in the launder-ometer and processed for 55 minutes (15 minutes heating time 22°C-60°C, 40 minutes holding time). The results obtained are shown in Table 11.

Table 11

Dosage-response with respect to PPT in launder-ometer scale.
 10 Conditions: 300 ml 0.02 M $(\text{NH}_4)_2\text{SO}_4/\text{NaHSO}_4$ pH 5.4 was added to a 1200 ml beaker together with 12 g denim (in one piece), 30 LACU TvL (to 0.1 LACU/ml), and PPT in the range 50 μM - 500 μM . The beakers were sealed and placed in the launder-ometer and processed for 55 minutes (15 minutes heating time 22°C-60°C, 40
 15 minutes holding time).

PPT (μM)	ΔL^*	Final pH
50 ~ 1.3 $\mu\text{mole/g}$	4.41	7.08
100 ~ 2.6 $\mu\text{mole/g}$	7.06	6.88
20 250 ~ 6.3 $\mu\text{mole/g}$	14.63	6.88
500 ~ 12.5 $\mu\text{mole/g}$	19.21	6.67

It can be seen from Table 11 that at the above given conditions the degree of bleaching is increased with the
 25 increased concentration of PPT.

EXAMPLE 10

Bleaching Denim with Peroxidase and Phenothiazine-10-Propionic Acid (PPT) using Different Buffers

To illustrate the bleaching process using peroxidase comparison of 2 high-performance buffers was made using PPT as an enhancing agent.

Method: Each buffer was prepared at a concentration of 0.01 M, and pH adjusted to pH 6.5 with NaOH or with the corresponding acid. 80 ml of the buffer in question was added to a 200 ml glass beaker together with a magnet bar (4 cm), and 8 circular pieces of denim (3.5 cm in diameter ~ 0.4 g), giving a denim:liquor ratio of 1:25. The glass beaker was incubated on a magnet stirrer (300 rpm) in a water bath at 50°C, and a pH electrode was dipped into the liquor in the middle of the beaker in order to monitor and control pH at pH 6.5 (i.e. the experiments were run under pH-stat conditions using a Radiometer pH-stat (PHM 82 or PHM 62 pH meter, TTT 80 Titrator, ABU 80 Autoburette) with automatic titration with the corresponding acid (0.1 M) if and when pH increased above pH 6.5). Following equilibration at pH 6.5, PPT (0.02 M in 96% ethanol) was added to a final concentration of 250 μM ~ 6.3 $\mu\text{mole/g}$ together with peroxidase from Coprinus cinereus (CiP, available from Novo Nordisk A/S) to a final concentration of 1 PODU/ml ~ 5 $\mu\text{g/g}$. Reaction was started adding 0.1 ml H_2O_2 (0.1 M) corresponding to a final concentration of 0.125 mM H_2O_2 . The concentration of H_2O_2 was monitored using peroxide sticks (Merckoquant Peroxid-Test, Merck. art. 10011). When the sticks indicated that the concentration of H_2O_2 was below 2 mg/l (0.059 mM), another 0.1 ml of H_2O_2 was added. However, the PPT radical seemed to interfere with the measurement in that the PPT radical itself (without the presence of H_2O_2) was able to colourize the sticks. But, for this purpose, only an indication of low concentration of H_2O_2 and low concentration of PPT radical was of interest, since no addition of hydrogen peroxide was needed even if the concentration was zero, as long as PPT radical was present. Only if the concentration of both H_2O_2 and

PPT radical was low/zero, more H_2O_2 should be added. After 30 minutes the denim swatches were rinsed in tap water and air dried on filter paper overnight, and the resulting degree of bleaching was determined as mentioned above in Example 1. The results are shown in Table 12 below.

Table 12

Comparison of bleach performance in acetate and oxalate buffer using 250 μM PPT and 1 PODU/ml peroxidase (CiP) at pH 6.5 and 50°C. H_2O_2 was added semi-continuously over time adding aliquots of 0.1 ml of a stock solution of 0.1 M H_2O_2 .

Acetate			Oxalate		
Time (minutes)	Conc. H_2O_2 (mg/l)	H_2O_2 added (ml)	Time (minutes)	Conc. H_2O_2 (mg/l)	H_2O_2 added (ml)
0	-	0.1	0	-	0.1
1	5	-	1	5	-
2.5	2	-	3	2	0.1
3	-	0.1	5	2	-
6	2	0.1	5.5	-	0.1
8	2	0.1	8	2	0.1
10.5	2	-	11	2	0.1
11	-	0.1	13	2	0.1
14	2	-	15	2	0.1
14.5	-	0.1	17	3.5	-
17	2	0.1	20	2	0.1
21	3.5	-	25	2	0.1
23	2	0.1	28	5	-
26	5	-	30	3	-
30	2.5	-	-	-	-
Total:	-	0.8	Total:	-	0.9
ΔL^* :	-	14.5	ΔL^* :	-	16.5

It can be seen from Table 12 that a high degree of bleaching can be achieved when using peroxidase, PPT and acetate or oxalate as buffers.

EXAMPLE 11

5 Large Scale Trials (40 Litres)

Large-scale trials (40 litres) with bleaching of denim using laccase and phenothiazine-10-propionic acid (PPT) have been performed giving the results shown below:

105.32 g $(\text{NH}_4)_2\text{SO}_4$, 25.48 g $\text{NaHSO}_4 \times 1 \text{ H}_2\text{O}$, 2.7 g PPT and
10 1.6 kg stonewashed denim was loaded into a wascator, giving a denim:liquor ratio of 1:25 and 6.3 $\mu\text{mole/g}$ denim of PPT.

40 litres of cold tap water was added together with
4000 LACU - 62500 μg Trametes villosa laccase (TvL - available
from Novo Nordisk A/S), giving 0.1 LACU/ml or 39 $\mu\text{g/g}$ denim of
15 TvL; temperature was raised to and maintained at 60°C for a
total processing time of 60 minutes, followed by a
rinsing/inactivation step with Na_2CO_3 (1 g/l) at 80°C for 15
minutes, followed by a rinse with tap water. After bleaching,
the denim was dried in a conventional tumble dryer.

20 The process resulted in a bleach level of $\Delta L^* = 16-17$.

EXAMPLE 12

Absorbed Organic Halogens (AOX)

As a result of the chlorine-free bleaching process, AOX was expected to be significantly lower using the enzymatic
25 approach compared to the conventional hypochlorite based
process. In Table 13 below, AOX data are shown for various
bleach levels using the enzymatic approach and the conventional
approach.

Table 13

30 Comparison of the resulting AOX-values of the processing liquor
following denim bleaching using conventional hypochlorite
process, and the enzymatic process. All experiments were run in

wascator scale (20-40 litres). The AOX values have been determined by VKI (the Danish Water Quality Institute).

Bleach Method	Bleach Level ΔL^*	AOX-values ppm
Hypochlorite	11.1	14
Hypochlorite	18.7	21
Peroxidase/PPT	5.6	<0.0025 ¹⁾
Laccase/PPT	17	n.d. ²⁾

¹⁾ Below detection limit

²⁾ Not determined

EXAMPLE 13

Strength Loss

The enzyme/enhancing agent bleaching process of the present invention results in a very specific attack on indigo and does not result in a damage of the cotton. This is illustrated in the strength loss of the processed denim. Using the enzyme/enhancing agent bleaching process the strength loss is much lower than by using the conventional hypochlorite process, which is illustrated in Table 14 below.

Stone washed denim was bleached to the same level using hypochlorite and using laccase/PPT, and the strength loss (tear strength of processed denim compared to tear strength of non-processed denim) determined. The results are shown in Table 14 below.

Table 14

Comparison of tensile strength loss using hypochlorite and using Laccase/PPT for bleaching of denim.

	ΔL^*	% tensile strength loss (warp)
NaOCl	17.99	15.8 %
Laccase/PPT	18.27	1.7 %

EXAMPLE 14Large-scale trials

5 Large-scale trials with bleaching of denim using laccase and phenothiazine-10-propionic acid (PPT) have been performed in a fulling machine (stainless steel drum of 1 m diameter and 0.4 m in dept operated at approx. 14 rpm) giving the results shown below:

10 The denim (75x100 cm) was sewn into "legs" (denim cylinders) weighing approximately 350-375 g each (not stone washed). 4 stone washed denim "legs" weighing 1458 g total, 40.8 g Na₂-oxalate, 12.0 g oxalic acid x 2 H₂O and 1.82 g PPT was loaded into the fulling machine, and 20 litres of hot
15 (55°C) tap water was added resulting in a pH of 5.5 increasing to 7.2 in 5 minutes. The liquor was heated to 60°C and pH was adjusted to pH 5.6 with 2 ml of 72% H₂SO₄, and 1824 LACU = 28500 µg TvL (Trametes villosa laccase, available from Novo Nordisk A/S) was added.

20 The conditions used were: 0.02 M oxalate buffer, pH 5.6-6.0, denim:liquor ratio = 1:14, 336 µM PPT = 4.6 µmole PPT/g denim, 0.09 LACU/ml = 19.5 µg enzyme protein/g denim. The bleaching was stopped after 30 minutes, and the denim rinsed with 2x20 litres of hot (55°C) tap water for 1-2
25 minutes. Following bleaching, the denim was dried in a conventional tumble drier.

The process resulted in a bleach level of ΔL^* : 17-18.

EXAMPLE 15Large-scale trials

Denim (75x100 cm) was sewn into "legs" (denim cylinders) weighing approximately 350-375 g each (not stone washed). 4 stone washed denim "legs" weighing 1480 g total, 24.2 g Na₂-oxalate, 12.5 g oxalic acid x 2 H₂O, and 1.75 g PPT was loaded into the fulling machine, and 14 litres of hot (55°C) tap water was added and heated to 60°C resulting in a pH of 4.8. pH was adjusted to pH 5.7 with 1.5 ml 50% NaOH. 1755 LACU = 27422 µg TvL was added (TvL = Trametes villosa laccase, available from Novo Nordisk A/S).

The conditions used were: 0.02 M oxalate buffer, pH 5.7-5.9, denim:liquor ratio = 1:10, 461 µM PPT = 4.4 µmole PPT/g denim, 0.13 LACU/ml = 18.5 µg enzyme protein/g denim. The bleaching was stopped after 30 minutes, and the denim rinsed with 2x40 litres of hot (55°C) tap water for 1-2 minutes. After bleaching, the denim was dried in a conventional tumble drier. The process resulted in a bleach level of ΔL*: 14-15.

EXAMPLE 16Industrial scale trials (450 litres)

Industrial scale trials (450 litres) with bleaching of denim using laccase and phenothiazine-10-propionic acid (PPT) have been carried out in a front loaded wash extractor (type: "Cherry Tree"), using 50 kg of denim (60 pair of jeans) and 450 litres of water, giving a denim:liquor (w/w) ratio of 1:9. Conditions and dosages were as follows:

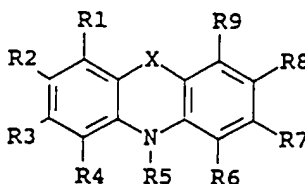
Trial 1: 450 g sodium phosphate
125 g di-sodium phosphate
125 g PPT ~ 9.2 µg/g denim
90.000 LACU (= 1406 mg) Trametes villosa laccase,
available from Novo Nordisk A/S ~ 29.2 µg/g denim
30 minutes, pH=6.2, 60°C

Trial 2: 1000 g di-sodium oxalate
175 g oxalic acid
125 g PPT ~ 9.2 µg/g denim
90.000 LACU (= 1406 mg) Trametes villosa laccase ~
s 29.2 µg/g denim
30 minutes, pH=5.5, 60°C

Following bleaching, the denim was tumble dried. The process resulted in bleach levels of $\Delta L^* = 16$ (trial 1) and $\Delta L^* = 21$ (trial 2).

CLAIMS

1. A process for providing a bleached look in the colour density of the surface of dyed fabric, the process comprising contacting, in an aqueous medium, a dyed fabric with a phenol oxidizing enzyme system and an enhancing agent of the following formula:



in which formula X represents (-O-) or (-S-), and the substituent groups R¹-R⁹, which may be identical or different, independently represents any of the following radicals:
 10 hydrogen, halogen, hydroxy, formyl, carboxy, and esters and salts hereof, carbamoyl, sulfo, and esters and salts hereof, sulfamoyl, nitro, amino, phenyl, C₁-C₁₄-alkyl, C₁-C₅-alkoxy, carbonyl-C₁-C₅-alkyl, aryl-C₁-C₅-alkyl; which carbamoyl, sulfamoyl, and amino groups may furthermore be unsubstituted or
 15 substituted once or twice with a substituent group R¹⁰; and which phenyl may furthermore be unsubstituted or substituted with one or more substituent groups R¹⁰; and which C₁-C₁₄-alkyl, C₁-C₅-alkoxy, carbonyl-C₁-C₅-alkyl, and aryl-C₁-C₅-alkyl groups may be saturated or unsaturated, branched or unbranched, and
 20 may furthermore be unsubstituted or substituted with one or more substituent groups R¹⁰;

which substituent group R¹⁰ represents any of the following radicals: halogen, hydroxy, formyl, carboxy and esters and salts hereof, carbamoyl, sulfo and esters and salts
 25 hereof, sulfamoyl, nitro, amino, phenyl, aminoalkyl, piperidino, piperazinyl, pyrrolidino, C₁-C₅-alkyl, C₁-C₅-alkoxy; which carbamoyl, sulfamoyl, and amino groups may furthermore be unsubstituted or substituted once or twice with hydroxy, C₁-C₅-alkyl, C₁-C₅-alkoxy; and which phenyl may furthermore be
 30 substituted with one or more of the following radicals: halogen, hydroxy, amino, formyl, carboxy and esters and salts

hereof, carbamoyl, sulfo and esters and salts hereof, and sulfamoyl; and which C₁-C₃-alkyl, and C₁-C₃-alkoxy groups may furthermore be saturated or unsaturated, branched or unbranched, and may furthermore be substituted once or twice with any of the following radicals: halogen, hydroxy, amino, formyl, carboxy and esters and salts hereof, carbamoyl, sulfo and esters and salts hereof, and sulfamoyl;

or in which general formula two of the substituent groups R¹-R⁹ may together form a group -B-, in which B represents any of the following the groups: (-CHR¹⁰-N=N-), (-CH=CH-)_n, (-CH=N-)_n or (-N=CR¹⁰-NR¹¹-), in which groups n represents an integer of from 1 to 3, R¹⁰ is a substituent group as defined above and R¹¹ is defined as R¹⁰.

2. A process according to claim 1, wherein the fabric is dyed with a vat dye such as indigo or thioindigo.

3. A process according to claim 1 or 2, wherein the fabric is a cellulosic fabric or a mixture of cellulosic fibres or a mixture of cellulosic fibres and synthetic fibres.

4. A process according to any of claims 1-3, wherein the fabric is denim, preferably denim dyed with indigo or thioindigo.

5. A process according to claim 1, in which the phenol oxidizing enzyme system is a peroxidase and a hydrogen peroxide source.

6. A process according to claim 5, wherein the peroxidase is horseradish peroxidase, soybean peroxidase or a peroxidase enzyme derived from Coprinus, e.g. C. cinereus or C. macrorhizus, or from Bacillus, e.g. B. pumilus, or Myxococcus, e.g. M. virescens.

7. A method according to claim 5 or 6, wherein the hydrogen peroxide source is hydrogen peroxide or a hydrogen peroxide precursor, e.g. perborate or percarbonate, or a

hydrogen peroxide generating enzyme system, e.g. an oxidase and its substrate, or a peroxycarboxylic acid or a salt thereof.

8. A method according to claims 1-7, wherein the aqueous medium contains H_2O_2 or a precursor for H_2O_2 in a concentration corresponding to 0.001-25 mM H_2O_2 .

9. A process according to claim 1, in which the phenol oxidizing enzyme system is a laccase or a laccase related enzyme together with oxygen.

10. A process according to claim 9, wherein the laccase is derived from Trametes, e.g., Trametes villosa, or Myceliophthora, e.g., Myceliophthora thermophila, or Coprinus, e.g., Coprinus cinereus.

11. A process according to claims 1-10, wherein the concentration of the phenol oxidizing enzyme corresponds to 0.001-10000 μ g of enzyme protein per g of denim.

12. A process according to claims 1-11, wherein the enhancing agent belongs to the group consisting of phenoxazine-10-propionic acid, phenoxazine-10-hydroxyethyl, phenothiazine-10-ethyl-4-carboxy, phenothiazine-10-propionic acid, promazine hydrochloride and phenothiazine-10-ethylalcohol.

13. A process according to claims 1-12, wherein the enhancing agent in the aqueous medium is present in concentrations of from 0.005 to 1000 μ mole/g denim.

14. A process according to claims 1-13 resulting in reduced strength loss of fabric compared to conventional bleaching processes using e.g. hypochlorite.

15. A process according to claims 1-14 resulting in zero AOX values or AOX values below detection limit in the processing liquor.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/DK 95/00418

A. CLASSIFICATION OF SUBJECT MATTER		
IPC6: D06P 3/02, C11D 3/386 According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED		
Minimum documentation searched (classification system followed by classification symbols)		
IPC6: D06P, C11D		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
SE,DK,FI,NO classes as above		
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)		
WPI, IFIPAT, CA		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 9412621 A (NOVO NORDISK A/S), 9 June 1994 (09.06.94), pages 15-16 in particular --	1-15
X	WO 9412619 A (NOVO NORDISK A/S), 9 June 1994 (09.06.94), pages 5-6 in particular --	1-11
A	WO 9218683 A (NOVO NORDISK A/S), 29 October 1992 (29.10.92) --	1-15
A	WO 9105839 A (NOVO NORDISK A/S), 2 May 1991 (02.05.91) -- -----	1-15
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input checked="" type="checkbox"/> See patent family annex.		
* Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family		
Date of the actual completion of the international search		Date of mailing of the international search report
14 February 1996		19-02-96
Name and mailing address of the ISA/ Swedish Patent Office Box 5055, S-102 42 STOCKHOLM Facsimile No. +46 8 666 02 86		Authorized officer Gerd Strandell Telephone No. +46 8 782 25 00

Form PCT/ISA/210 (second sheet) (July 1992)

INTERNATIONAL SEARCH REPORT

05/01/96

International application No.

PCT/DK 95/00418

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
WO-A-	9412621	09/06/94	NONE	
WO-A-	9412619	09/06/94	NONE	
WO-A-	9218683	29/10/92	NONE	
WO-A-	9105839	02/05/92	AT-T- 108484	15/07/94
			AU-B- 646645	03/03/94
			AU-A- 6515790	16/05/91
			AU-A- 6516090	16/05/91
			CA-A- 2067748	14/04/91
			CN-A- 1051600	22/05/91
			DE-D,T- 69010691	16/03/95
			EP-A,B- 0495836	29/07/92
			SE-T3- 0495836	
			EP-A,B- 0497794	12/08/92
			SE-T3- 0497794	
			ES-T- 2057593	16/10/94
			JP-T- 5500899	25/02/93
			JP-T- 5503542	10/06/93
			TR-A- 26687	15/05/95
			US-A- 5273896	28/12/93
			WO-A- 9105858	02/05/91

Form PCT/ISA/210 (patent family annex) (July 1992)